ORIGINAL ARTICLE



Structures of New Cytotoxic Antibiotics, Piericidins C7 and C8

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Abstract Two new cytotoxic antibiotics, piericidins C_7 (1) and C_8 (2), were isolated from a marine *Streptomyces* sp. High-resolution FAB-MS established the molecular formulae of 1 and 2 as $C_{28}H_{41}NO_5$ and $C_{29}H_{43}NO_5$, respectively. The planar structures of the piericidins were elucidated by NMR spectral analysis including COSY, HMQC and HMBC. The relative stereochemistry of 2 was determined based on NOESY and coupling constant analyses. 1 and 2 are new members of the piericidin family possessing 11,12-epoxide and 12-butenyl groups.

Keywords piericidin, cytotoxic antibiotic, epoxide

Introduction

In the course of screening for antitumor antibiotics against cells transformed with the adenovirus E1A gene, two new active substances belonging to the piericidin family were isolated from the fermentation broth of a marine *Streptomyces* sp. and were named piericidins C_7 (1) and C_8 (2). In the preceding paper [1], we have described the fermentation, isolation, physico-chemical properties and biological activities of 1 and 2, as well as the taxonomy of the producing organism. This paper describes the structure elucidation of 1 and 2.

Results and Discussion

Structure Elucidation

High-resolution FAB-MS established the molecular formulae of **1** and **2** as $C_{28}H_{41}NO_5$ (*m/z* 472.3062 [MH]⁺, Δ -0.1 mmu) and $C_{29}H_{43}NO_5$ (*m/z* 486.3223 [MH]⁺, Δ +0.4 mmu), respectively. The IR spectra of **1** and **2** revealed the presence of hydroxyl groups (3400 cm⁻¹) and the absence of carbonyl groups. ¹³C and ¹H NMR data for **1** and **2** are summarized in Table 1. All one-bond ¹H-¹³C connectivities were confirmed by a heteronuclear multiplequantum coherence (HMQC) experiment [2].

Three separate proton spin systems in 2 depicted in Fig. 1 by bold lines were identified by a COSY experiment and were expanded to a 3,5,7,9,11,13-hexamethylpentadeca-2,5,7,13-tetraenyl moiety based on ¹H-¹³C long-range couplings from five singlet methyls (3-Me, 5-Me, 7-Me, 11-Me and 13-Me) to the relevant carbons in the heteronuclear multiple-bond correlation (HMBC) [3] spectrum (Fig. 1). Oxygen substituents were located on C-10, C-11 and C-12 from their chemical shifts (δ 82.7, 66.0 and 66.0, respectively). The remaining five aromatic carbons and a nitrogen atom were required to construct a pyridine moiety. The substitution of the pyridine ring was determined by ¹H-¹³C long-range correlations from 1-H to C-1' and C-2', from a singlet methyl ($\delta_{\rm H}$ 2.03) to C-1', C-2' and C-3', from a methoxyl ($\delta_{\rm H}$ 3.72) to C-4' and from a methoxyl ($\delta_{\rm H}$ 3.89) to C-5'. A low-field ¹³C chemical shift $(\delta 151.5)$ for C-3' indicated the presence of a hydroxyl substituent on C-3'. The existence of an epoxide ring between C-11 and C-12 was required by their characteristic ¹³C chemical shifts (δ 66.0×2) and the molecular formula of 2. The geometrical configurations were elucidated to be 2E, 5E, 7E and 13E from high-field 13 C chemical shifts for five allylic methyls and NOESY correlations between 2-H

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and $4-H_2$, between $4-H_2$ and 6-H, and between 6-H and 8-H (Fig. 2).

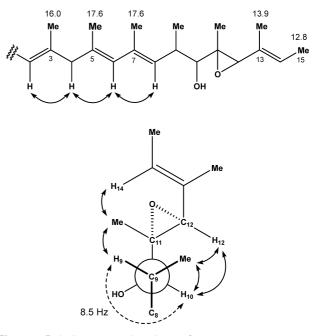
The relative configurations of **2** were analyzed by NOE correlations and coupling constants. The epoxide stereochemistry was identified as *trans* by NOEs between 10-H and 12-H and between 11-Me and 14-H. A large

 Table 1
 NMR data summary for 1 and 2 in CD₃OD

No	1		2	
	$\delta_{ ext{C}}$	$\delta_{ m H}$ multiplicity (Hz)	$\delta_{ ext{C}}$	$\delta_{ m H}$ multiplicity (Hz)
1	35.2	3.34 2H d (7.0)	35.2	3.35 2H d (7.0)
2	123.4	5.30 t (7.0)	124.7	5.30 t (7.0)
3	135.9		135.1	
4	43.9	2.75 2H d (7.0)	51.9	2.69 2H s
5	126.7	5.55 dt (15.5, 7.0)	135.0	
6	137.5	6.06 d (15.5)	131.7	5.66 s
7	134.9		134.3	
8	134.7	5.31 d (10.0)	133.3	5.17 d (10.0)
9	37.7	2.68 m	37.7	2.62 m
10	82.6	2.99 d (8.5)	82.7	2.97 d (8.5)
11	65.9		66.0	
12	65.8	3.23 s	66.0	3.25 s
13	130.6		130.5	
14	121.8	5.41 q (7.0)	121.8	5.42 q (7.0)
15	12.8	1.64 3H d (7.0)	12.8	1.64 3H d (7.0)
1′	151.6		151.5	
2′	114.5		114.6	
3′	151.6		151.5	
4′	130.1		130.1	
5′	156.0		156.0	
3-Me	16.7	1.72 3H s	16.0	1.67 3H s
5-Me			17.6	1.65 3H s
7-Me	13.1	1.74 3H s	17.6	1.72 3H s
9-Me	18.2	0.98 3H d (7.0)	18.2	0.99 3H d (7.0)
11-Me	10.6	1.09 3H s	10.6	1.11 3H s
13-Me	13.9	1.65 3H s	13.9	1.65 3H s
2'-Me	10.8	2.03 3H s	10.9	2.03 3H s
4'-OMe	60.8	3.72 3H s	60.9	3.72 3H s
5'-OMe	53.8	3.89 3H s	53.9	3.89 3H s

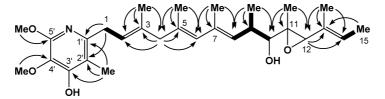
coupling constant (8.5 Hz) revealed an anti relationship between 9-H and 10-H. NOEs from 9-Me to 10-H and 12-H required these atoms to exist on the same side of a plane representing C-8, C-9, C-10 and C-11 as shown in Fig. 2. An NOE between 9-H and 11-Me arranged them on the opposite side (Fig. 2), thereby establishing the structure of 2 including the relative stereochemistry as shown in Fig. 3.

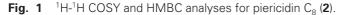
The ¹³C and ¹H NMR spectra of **1** showed a loss of one methyl in comparison with **2** (Table 1). A newly appeared olefinic proton (δ 5.55) was coupled with a methylene (4-H₂) and an olefinic methine (6-H), and its *E* geometry was determined by a large vicinal coupling constant (15.5 Hz). The remaining spectral features were essentially identical with those of **2**, indicating that **1** is a 5-demethyl derivative of **2**. The structure of **1** (Fig. 3) was confirmed by COSY, HMQC, HMBC and NOESY experiments (data not shown).





Solid arrows show NOE correlations and dashed arrows indicate $^1\mbox{H-}^1\mbox{H}$ couplings.





Bold lines show ¹H spin networks and arrows indicate ¹H-¹³C long-range correlations.

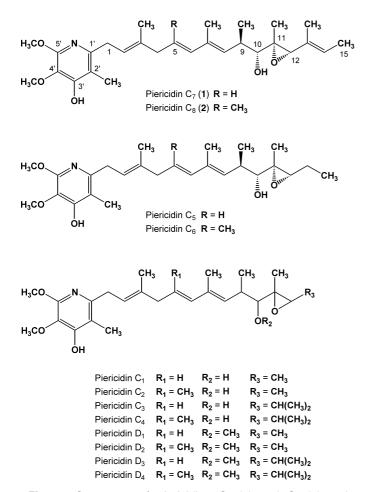


Fig. 3 Structures of piericidins C_7 (1) and C_8 (2) and related compounds.

A series of piericidins possessing an epoxide moiety have been isolated from *Streptomyces* sp. and *Nocardioides* sp., *e.g.*, piericidins $C_1 \sim C_6$ and piericidins $D_1 \sim D_4$ (Fig. 3) [4~7]. 1 and 2 represent two new members of this family and are the first examples produced by a marine actinomycete.

Experimental

Spectroscopic Measurements

Mass spectra were obtained on a JEOL SX-102A

spectrometer in the FAB mode using *m*-nitrobenzyl alcohol as matrix and polyethylene glycol as internal standard. IR spectra were measured on a JASCO FT/IR-410 spectrometer. NMR spectra were obtained on a JEOL JNM-LA400 spectrometer with ¹H NMR at 400 MHz and with ¹³C NMR at 100 MHz. Chemical shifts are given in ppm relative to CHD₂OD at 3.30 ppm for ¹H NMR and CD₃OD at 49.0 ppm for ¹³C NMR.

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